



A nuclear single-nucleotide polymorphism (SNP) potentially useful for the separation of *Rhodnius prolixus* from members of the *Rhodnius robustus* cryptic species complex (Hemiptera: Reduviidae)

Márcio G. Pavan^a, Rafael D. Mesquita^b, Gena G. Lawrence^c, Cristiano Lazoski^d, Ellen M. Dotson^c, Sahar Abubucker^e, Makedonka Mitreva^e, Jennifer Randall-Maher^e, Fernando A. Monteiro^{a,*}

^a Laboratório de Sistemáticas Bioquímica, Instituto Oswaldo Cruz, FIOCRUZ, Rio de Janeiro, Brazil

^b Laboratório de Bioinformática, Departamento de Bioquímica, Instituto de Química, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

^c Centers for Disease Control and Prevention, Division of Parasitic Diseases and Malaria, Entomology Branch, 1600 Clifton Road NE, Atlanta, GA 30329, United States

^d Laboratório de Biodiversidade Molecular, Instituto de Biologia, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

^e The Genome Institute, Washington University School of Medicine, Campus Box 8501, St. Louis, MO 63108, United States

ARTICLE INFO

Article history:

Received 19 June 2012

Received in revised form 16 October 2012

Accepted 23 October 2012

Available online 6 December 2012

Keywords:

Rhodnius

Cryptic species

SNP

Nuclear DNA

ABSTRACT

The design and application of rational strategies that rely on accurate species identification are pivotal for effective vector control. When morphological identification of the target vector species is impractical, the use of molecular markers is required. Here we describe a non-coding, single-copy nuclear DNA fragment that contains a single-nucleotide polymorphism (SNP) with the potential to distinguish the important domestic Chagas disease vector, *Rhodnius prolixus*, from members of the four sylvatic *Rhodnius robustus* cryptic species complex. A total of 96 primer pairs obtained from whole genome shotgun sequencing of the *R. prolixus* genome (12,626 random reads) were tested on 43 *R. prolixus* and *R. robustus* s.l. samples. One of the seven amplicons selected (*AmpG*) presented a SNP, potentially diagnostic for *R. prolixus*, on the 280th site. The diagnostic nature of this SNP was then confirmed based on the analysis of 154 *R. prolixus* and *R. robustus* s.l. samples representing the widest possible geographic coverage. The results of a 60% majority-rule Bayesian consensus tree and a median-joining network constructed based on the genetic variability observed reveal the paraphyletic nature of the *R. robustus* species complex, with respect to *R. prolixus*. The *AmpG* region is located in the fourth intron of the *Transmembrane protein 165* gene, which seems to be in the *R. prolixus* X chromosome. Other possible chromosomal locations of the *AmpG* region in the *R. prolixus* genome are also presented and discussed.

© 2013 Elsevier B.V. All rights reserved.

1. Introduction

Chagas disease is transmitted through *Trypanosoma cruzi* infected feces of triatomine bugs (Hemiptera: Reduviidae) and is endemic in Latin America and in the Caribbean (Coura, 2007), ranking fourth in epidemiologic importance among all neglected diseases in those regions (Hotez et al., 2008). Although case incidence estimates show a 90% decline (from 500,000 to 50,000 new cases per year), 8 million people are still infected and 14,000 die every year (Senior, 2007). Domestic insect vectors are responsible for most of the disease transmission to humans. Because no vaccine is available, the elimination of household-infesting triatomines is the

major goal of many Chagas disease control programs (Abad-Franch et al., 2010).

The design and implementation of any vector control intervention must rely on accurate species identification. However, the identification of Chagas disease vectors based on morphological characters alone can be technically challenging, since some groups of species are cryptic (i.e., isomorphic), and therefore not amenable to be distinguished in such a manner. This taxonomical issue is even more relevant when applied to the transmission of *T. cruzi* by *Rhodnius prolixus* and *Rhodnius robustus* s.l. to humans. Although morphologically indistinguishable, they play very different epidemiological roles: the former are efficient domestic vectors, and the latter are not, as all species of the complex are entirely sylvatic (Monteiro et al., 2003).

Rhodnius prolixus is one of the most important Chagas disease vectors in Latin America, particularly in Venezuela, Colombia, and Central America, due to its ability to efficiently colonize human dwellings. Although certain countries such as Guatemala have successfully controlled the transmission mediated by this species

* Corresponding author. Address: Laboratório de Sistemáticas Bioquímica, Instituto Oswaldo Cruz, FIOCRUZ, Avenida Brasil, 4365, Pavilhão Arthur Neiva, sala 21A/22, Manginhos, Rio de Janeiro, RJ 21045-900, Brazil. Tel.: +55 21 25621255; fax: +55 21 22803740.

E-mail address: fam@ioc.fiocruz.br (F.A. Monteiro).

(Guhl et al., 2009), Venezuela and Colombia still face serious problems of re-infestation of insecticide-treated houses by sylvatic populations (Fitzpatrick et al., 2008; Lopez et al., 2007). These events are commonplace in endemic regions where vectors are autochthonous, and will demand the development of innovative surveillance/control strategies that contemplate the complexities of sylvatic populations dynamics (Abad-Franch and Monteiro, 2005).

To allow for the accurate taxonomic identification of *R. prolixus* and the members of the *R. robustus* cryptic species complex, Pavan and Monteiro (2007) have developed an objective and cost-effective mitochondrial DNA (mtDNA) multiplex-PCR assay. However, due to its mitochondrial-based nature, this method could generate misleading results in areas where *R. prolixus* and members of the *R. robustus* complex might come into contact and hybridize, causing mitochondrial DNA introgression (Fitzpatrick et al., 2008).

Thus, in order to overcome such limitation, we describe a new non-coding, single-copy nuclear DNA fragment containing a single-nucleotide polymorphism (SNP) with the potential to separate *R. prolixus* from members of the *R. robustus* cryptic species complex.

2. Materials and methods

2.1. Molecular identification of specimens and sampling

Since accurate taxonomic identification of *R. prolixus* and the four cryptic species of *R. robustus* [sensu Monteiro et al. (2003)] based on morphology alone is problematic, all specimens used in this study were identified through molecular taxonomy based on a fragment of the mitochondrial cytochrome *b* gene (Pavan and Monteiro, 2007). Moreover, we sequenced the ribosomal second internal transcribed spacer (ITS-2) of all specimens analyzed (data not shown), according to Marcilla et al. (2001), to identify possible introgression of mitochondrial DNA specimens.

For an exploratory screening aimed at finding unique, non-coding regions that separate these species, we tested 25 *R. prolixus* from two well established laboratory colonies kept at the Centers for Disease Control and Prevention (CDC) in Atlanta (19 specimens) and at the Medical Entomology Research and Training Unit in Guatemala (six specimens), plus two field-collected *R. prolixus* from Venezuela, two *R. robustus* I, four of each *R. robustus* II and III, and six *R. robustus* IV [sensu Monteiro et al. (2003)]. This approach was done during the initial phase of *Rhodnius prolixus* genome project, named *survey sequencing*, aimed to select a non-introgressed *R. prolixus* colony with low genetic variability. Fifty-two colony specimens, 43 *R. prolixus* and nine *R. robustus* II, and 102 field-collected insects, three *R. prolixus*, two *R. robustus* I, 74 *R. robustus* II, 19 *R. robustus* III, and four *R. robustus* IV (Fig. 1 and Table 1), were

assayed to confirm the diagnostic applicability of possible species-specific SNPs.

2.2. DNA purification and library construction

DNA from the CDC and Guatemala Colony samples were extracted from ovaries or testis using the Promega genomic extraction kit. For all other *Rhodnius* specimens, one or two legs of a single *Rhodnius* specimen was placed in a 1.5 mL microtube, dipped into liquid nitrogen until frozen, and ground to a powder with Kimble/Kontes Pellet Pestles® prior to standard Genomic DNA Extraction with a Real Genomics™ kit.

From a small WGS plasmid library, 12,626 random Genome Survey Sequences (GSS) were generated as described by Abubucker et al. (2008), using genomic DNA extracted from a population of *R. prolixus* from the Atlanta colony, and running on an ABI 3730 automated sequencer. Ten thousand GSS reads were screened for non-coding regions, searched against a transposon database, then against RFAM (Gardner et al., 2011), and finally against a non-redundant protein database (with an *e*-value cutoff of 10^{-5}). Regions with hits were masked. Vector sequences were removed and sequences with unmasked regions of 400–500 base pairs (bp) were extracted and sent through an in-house primer-calling pipeline. Primers with the following criteria were picked: (1) all bases of each primer in the pair must have a quality score of at least Q25; (2) minimum and maximum acceptable amplicon size was 400 and 500, respectively; and (3) excluded regions were 150-bp long (i.e., region with repetitive elements in the middle of the amplicon that would compromise primer annealing).

2.3. PCR amplification and DNA sequencing

From the putative unique, non-coding regions, 96 primer pairs were designed for the PCR amplification of regions of approximately 400–500-bp in length (Table A.1), during the *survey sequencing* approach.

PCR amplifications of the nuclear gene fragments were performed in a Veriti® 96 Well Thermal Cycler (Applied Biosystems), programmed for one denaturation step at 96 °C for 5 min, followed by 35 cycles at 94 °C for 30 s, 60 °C for 30 s, and 72 °C for 45 s, and a final 10-min extension step at 72 °C. Each 25 µL final volume reaction, contained 5–10 ng of DNA template, 2.5 µL buffer 10×, 2.5 mM of MgCl₂, 1.25 units of Taq polymerase (Biotools), 0.5 mM of dNTPs, and 10 pmol of each primer. Purification of PCR products was performed with the Hi Yield™ Gel/PCR DNA Extraction Kit (Real Genomics™), and both fragment strands were subjected to fluorescent dye-terminator cycle sequencing reactions (ABI Prism® BigDye® Terminator v3.1 Cycle Sequencing Kit, Applied Biosystems), using the same primers above, and run on an ABI 3730 automated sequencer.

2.4. Analyses of generated DNA sequence data

The removal of primer sequences, editing of both forward and reverse strands, and the generation of a consensus sequence for each sample were done using the SEQMAN LASERGENE 7.0 program (DNASTar, Inc.). Sequences were aligned using the default parameters of CLUSTALW2 program (Larkin et al., 2007). Polymorphic sites were identified using MEGA 5 (Tamura et al., 2011). RECON (Bao and Eddy, 2002), and BLASTX (Altschul et al., 1997) were used to identify repeated regions and coding sequences, respectively.

A Bayesian phylogenetic tree constructed based on the diagnostic amplicons for 154 *R. prolixus* and *R. robustus* s.l. sequences was inferred in BEAST 1.7 (Drummond et al., 2012). Bayesian Information Criterion (BIC) in jMODELTEST (Posada, 2008) was used to elect Jukes and Cantor (1969) as the best-fit evolutionary model (delta

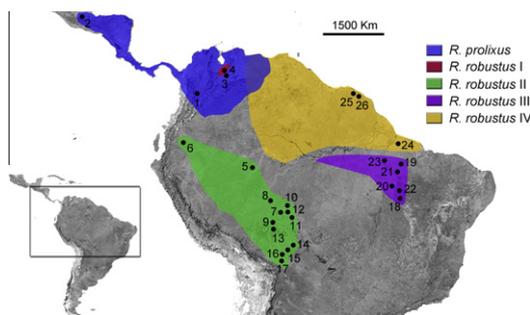


Fig. 1. Geographical distribution of sampled specimens of *R. prolixus* and *R. robustus* s.l. Each location is numbered according to Table 1. The colors indicate the range of *R. prolixus* and the four members of the *R. robustus* cryptic species complex (*R. robustus* I–IV).

Table 1
Information on the samples used in this study. Field-collected samples are numbered according to Fig. 1. N_A – number of individuals tested for seven amplicons (*AmpA* to *AmpG*); N_B – number of individuals sequenced for *AmpG*. ¹Centers for Disease Control and Prevention; ²Medical Entomology Research and Training Unit; ³Instituto Oswaldo Cruz; ⁴Instituto Nacional de Pesquisa da Amazonia.

Species	Collection site/colony location	Geographical coordinates	N _A	N _B	Field/colony	Date of collection	GenBank accession numbers	
<i>R. prolixus</i>	1. Colombia/CDC ¹ , Atlanta, USA	–	19	10	Colony	–	JQ432871 to 432880	
	2. Guatemala/MERTU ² , Guatemala	–	6	10	Colony	–	JQ432881 to 432890	
	“Unknown”/IOC ³ , Brazil	–	–	23	Colony	–	JQ432844 to 432866	
	3. Barinas, Venezuela	08°37'N 70°12'W	2	3	Field	2004	JQ432891 to 432893	
<i>R. robustus</i> I	4. Trujillo, Venezuela	09°22'N 70°25'W	2	2	Field	2004	JQ432894, JQ432895	
<i>R. robustus</i> II	5. Caruarí, Amazonas, Brazil/INPA ⁴ , Brazil	–	–	4	Colony	–	JQ432867 to 432870	
	6. Sucumbios, Ecuador	–	–	5	Colony	–	JQ432896 to 432900	
	7. Monte Negro, Rondônia, Brazil	10°15'N 63°17'W	4	2	Field	2004	JQ432961, JQ432962	
	8. Porto Velho, Rondônia, Brazil	08°44'S 63°25'W	–	45	Field	Apr/2009	JQ432916 to 432960	
	9. Cacoal, Rondônia, Brazil	11°26'S 61°27'W	–	2	Field	Apr/2009	JQ432973, JQ432974	
	10. Santo Antônio, Rondônia, Brazil	12°31'S 63°33'W	–	1	Field	Feb/2010	JQ432971	
	11. Guajará Mirim, Rondônia, Brazil	10°47'S 65°20'W	–	2	Field	Jul/2010	JQ432969, JQ432970	
	12. Ouro Preto do Oeste, Rondônia, Brazil	10°43'S 62°15'W	–	7	Field	Oct/2009	JQ432963 to 432968, JQ432972	
	13. San Gabriel, Pando, Bolivia	10°16'S 67°03'W	–	2	Field	Sep/2007	JQ432912, JQ432913	
	14. Guarayos, Santa Cruz, Bolivia	15°52'S 63°20'W	–	7	Field	Mar/2008	JQ432901 to 432907	
	15. San Ramón, Santa Cruz, Bolivia	16°29'S 62°29'W	–	1	Field	Mar/2008	JQ432908	
	16. Montero, Santa Cruz, Bolivia	17°17'S 63°08'W	–	3	Field	Mar/2008	JQ432909 to 432911	
	17. El Torno, Santa Cruz, Bolivia	18°04'S 63°18'W	–	2	Field	Sep/2007	JQ432914, JQ432915	
	<i>R. robustus</i> III	18. Araguaína, Pará, Brazil	07°11'S 48°12'W	–	2	Field	Nov/2009	JQ432975, JQ432976
		19. Ulianópolis, Pará, Brazil	03°43'S 47°29'W	–	7	Field	Nov/2009	JQ432977 to 432983
		20. São Domingos do Araguaia, Pará, Brazil	05°30'S 48°44'W	–	3	Field	Nov/2009	JQ432984 to 432986
		21. Rondon do Pará, Pará, Brazil	04°54'S 48°20'W	–	2	Field	Nov/2009	JQ432987, JQ432988
22. São Bento do Tocantins, Tocantins, Brazil		06°02'S 47°55'W	–	3	Field	Nov/2009	JQ432989 to 432991	
23. Novo Repartimento, Pará, Brazil		04°20'S 49°50'W	4	2	Field	Aug/1998	JQ432992, JQ432993	
<i>R. robustus</i> IV	24. Santa Maria, Pará, Brazil	01°20'S 47°32'W	–	1	Field	Oct/2009	JQ432994	
	25. Cayenne, French Guiana	04°56'N 52°20'W	5	2	Field	Jan/2003	JQ432995, JQ432996	
	26. Rémire, French Guiana	04°53'N 52°16'W	1	1	Field	Aug/2001	JQ432997	
Total			43	154				

AIC = 0) for the data set. Tree prior was randomly generated and Yule process of speciation was imposed for all tree reconstructions. Two independent runs were performed for 10⁷ generations, with a burn-in of 10⁶ generations. Convergence of parameters and a proper mixing were confirmed by calculating the Effective Sample Size (ESS) in TRACER 1.5 (Drummond and Rambaut, 2007), excluding the initial 10% (burn-in) of each run. All considered parameters showed ESS values above 10⁴. Runs were combined using LOGCOMBINER, and a maximum credibility tree based on the 20,000 trees generated (burn-in = 2000) and a posterior probability limit of 0.6 was produced using TREE ANNOTATOR (both programs part of the BEAST package). Statistical support for clades was assessed by the posterior probability method, and trees were visualized in FIGTREE v.1.0 (<http://tree.bio.ed.ac.uk/software/figtree/>).

Genealogy of the DNA sequences generated for the selected diagnostic amplicon was also inferred with NETWORK 4.6 (Fluxus-Engineering), using the Median-Joining network method (Bandelt et al., 1999), with a Maximum-Parsimony (MP) calculation in the post-processing. Character weights and epsilon values were given a value of 10 (transition/transversion weighting was 1:1, since we analyzed a non-coding region). Also the Maximum-Composite Likelihood (M-CL) algorithm (Tamura et al., 2004) with 1000 bootstrap replicates of variance, available in MEGA 5 (Tamura et al., 2011), was used to construct intra and interspecific distance matrices.

2.5. Genomic search of selected amplicon

Using the MEGABLAST web service (http://www.ncbi.nlm.nih.gov/sutils/genom_table.cgi), the diagnostic amplicon was searched for in the following 40 complete or partial arthropod genomes:

Acromyrmex echinaior, *Acyrtosiphon pisum*, *Aedes aegypti*, *Anopheles gambiae*, *Anopheles darlingi*, *Apis mellifera*, *Apis florea*, *Atta cephalotes*, *Bombus impatiens*, *Bombus terrestris*, *Bombyx mori*, *Camponotus floridanus*, *Culex quinquefasciatus*, *Daphnia pulex*, *Drosophila melanogaster*, *Drosophila pseudoobscura*, *Drosophila obscura*, *Drosophila simulans*, *Drosophila yakuba*, *Drosophila ananassae*, *Drosophila erecta*, *Drosophila grimshawi*, *Drosophila mojavensis*, *Drosophila persimilis*, *Drosophila sechellia*, *Drosophila virilis*, *Drosophila willistoni*, *Harpegnathos saltator*, *Ixodes scapularis*, *Lepeophtheirus salmonis*, *Mayetiola destructor*, *Nasonia giraulti*, *Nasonia longicornis*, *Pediculus humanus corporis*, *Pogonomyrmex barbatus*, *Rhipicephalus microplus*, *R. prolixus*, *Solenopsis invicta*, *Varroa destructor*, *Nasonia vitripennis*, and *Tribolium castaneum*. A second search was done in the BLASTN web service (http://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastn&BLAST_PROGRAMS=megaBlast&PAGE_TYPE=BlastSearch&SHOW_DEFAULTS=on&LINK_LOC=blasthome), with standard parameters of word size = 7 and *e*-value = 10, and the NCBI nucleotide (nt) database arthropod (taxid 6656) subset.

2.6. Selected amplicon genomic contig localization and chromosome synteny

2.6.1. Databases

The *R. prolixus* genome assembly v.3.0 and the hard-masked contig GL563069 (“N” masked transposons, repetitive and low complexity regions) were downloaded from VectorBase (<http://www.vectorbase.org>). Transposable elements and *Rhodnius* EST databases were kindly provided by Dr. J.M. Ribeiro. *A. mellifera*, *B. terrestris* and *T. castaneum* protein sequences were extracted from the NCBI non-redundant (*nr*) database using a Perl script that selected and extracted all sequences related to any NCBI taxonomic

index point (taxids used were respectively 7460, 30195 and 7070), using taxdump and gi-taxid-prot NCBI relational datasets (<ftp://ftp.ncbi.nih.gov/pub/taxonomy/taxdump.tar.gz>, and ftp://ftp.ncbi.nih.gov/pub/taxonomy/gi_taxid_prot.dmp.gz). The *T. castaneum* genome assembly version 3.0 was downloaded from BeetleBase (ftp://bioinformatics.ksu.edu/pub/BeetleBase/3.0/Tribolium_genome_sequence.fasta). *R. prolixus* genes, transcripts and peptides belonging to prediction version C1 (VectorBase gene prediction pipeline using assembly v3.0) were also downloaded from VectorBase (named VectorBase prediction). Alternative gene prediction version Lagerblad 3.0 was done using GENEID v1.3 (Blanco et al., 2007) trained with a large *R. prolixus* EST database in genome assembly v3.0 (named Lagerblad and available in *R. prolixus* genomic browser at VectorBase). Transposable elements were identified in contig GL563069 using local RPSBLAST, BLASTN and TBLASTX (Tatusova, 2010), and transposable element databases.

2.6.2. Selected amplicon localization in genomic contigs

The selected amplicon region was searched on *R. prolixus* genomic contigs using BLASTN (Tatusova, 2010).

2.6.3. Genes and transposons curation in amplicon flanking regions

Genes from both predictions flanking the diagnostic amplicon region were searched for chimeras using BLASTP, NCBI nr protein, UNIPROT databases, and TBLASTN of *Rhodnius* EST database. Chimera genes were split manually to allow gene synteny analysis. Gene prediction revision was done manually on ARTEMIS (Rutherford et al., 2000) based on EST database hits and GENEWISE (Birney et al., 2004) analysis using contig GL563069 and coded proteins best-hit in nr and UNIPROT databases. Transposable elements revision was also carried out in GENEWISE with BLASTN and TBLASTX hits, using TE database.

2.6.4. Gene synteny

Reciprocal BLASTP best-hit strategy (Tatusov et al., 1997) was used to identify homologs at the protein level for 14 genes in the diagnostic amplicon flanking region. We used in the comparison species with chromosome-mapped genomes (*A. mellifera*, *B. terrestris* and *T. castaneum*). The *R. prolixus* database included predicted proteins from both VectorBase and Lagerblad predictions. Bee and beetle homolog proteins GIs from the NCBI protein database (<http://www.ncbi.nlm.nih.gov/protein>) were used to identify coding genes and their chromosome location. Transposable elements were not considered in this analysis.

2.6.5. Whole contig synteny

Rhodnius prolixus hard-masked contig GL563069 was compared with all *T. castaneum* chromosomes separately using TBLASTX with a 10^{-10} *e*-value cutoff. The results were loaded in ACT (Rutherford et al., 2000) and limited by 50 or 70% identity cutoff when confirming selected gene homolog location, and 50% identity in GL563069 open reading frames (ORFs) larger than 100 bp when analyzing entire contig synteny.

3. Results

3.1. Amplicon selection

Seven amplicons were selected based on reproducibility, variability, and presence of species-specific sites across all taxa tested. In this first screening, 27 *R. prolixus*, two *R. robustus* I, four of each *R. robustus* II and III, and six *R. robustus* IV samples PCR-amplified and sequenced for these seven nuclear loci yielded 13 polymorphic sites, nine of which are SNPs (Table A.2). One such SNP located on the 280th site of the 364-bp region amplified with primers

AmpGF, 5'–GAG AGC TGA AGA TAG GCA AGC G, and *AmpGR*, 5'–TGA TAA CTG GAT TAG GCG CAG C was diagnostic for *R. prolixus* (i.e., all *R. prolixus* have an adenine, instead of a guanine, on that particular site). To further evaluate the reliability and usefulness of this particular SNP to serve as an important diagnostic marker for the identification of *R. prolixus*, an extensive effort was made to obtain the greatest amount of *R. prolixus* and *R. robustus s.l.* samples aimed at covering the widest geographical distribution for the species. A total of 154 specimens, 46 *R. prolixus*, two *R. robustus* I, 83 *R. robustus* II, 19 *R. robustus* III, and four *R. robustus* IV, were sequenced for the 364-bp *AmpG* region. The 280th site still proved to be diagnostic. This autapomorphic character of *R. prolixus* is preceded by 5'–GTCATAAGA and followed by 5'–TCGTTGCTAG conserved sequences for all species analyzed.

Ten polymorphic sites were observed, eight of which are shared between *R. prolixus* and *R. robustus* I, and three of those are parsimony-informative sites (positions 174, 270, and 360; Table 2). The paraphyletic assemblage of *R. robustus s.l.*, previously observed with mitochondrial cytochrome *b* and ribosomal DNA sequencing analyses (Monteiro et al., 2003), is corroborated here, based on a Bayesian phylogenetic tree reconstruction and a Median-Joining network with Maximum-Parsimony post-processing (Fig. 2). The *R. robustus* I clade is more closely related to the *R. prolixus* clade than to the other *R. robustus* clades. The *R. robustus* I and *R. prolixus* sister-species status is also evidenced by M-CL distance comparisons, as sequence divergence between these species is lower than between *R. robustus* I and *R. robustus* II–IV (0.6% in comparison with 1.1–1.7%; Table 3).

3.2. Amplicon G genomic contig and flanking region

The *AmpG* region is located in the *R. prolixus* genomic contig GL563069 from positions 1116977 to 1117340. It was not found in any other genome searched, not even when high *e*-values and smaller word sizes were used (allowing for more dissimilar matches to occur), which suggests that this region is unique.

Genes in the *AmpG* flanking region (contig GL563069 from 925995 to 1319022 positions) were curated (Fig. 3B) to remove chimera and wrong predictions. Lagerblad genes 17948_72 and 17948_73 were joined (*Src tyrosine kinase*) and 17948_78 was split into two genes (*X11Lbeta* and *Transmembrane protein 165*). Genes 17948_79 and 17948_84 were wrong predictions and then were removed (not shown). No chimera or false positive gene was found in the VectorBase prediction, but four genes were absent (17948_70 – *Ephrin*, 17948_75 – *Sarcoplasmic calcium-binding protein*, 17948_81 – *Interleukin 16* and 17948_88 – *KN motif and Ankyrin repeat domain-containing protein*). Final genes in the *AmpG* flanking regions were: *Ephrin*, *Ribonuclease*, *Src tyrosin kinase*, *Odorant receptor*, *Sarcoplasmic calcium-binding protein*, *Small glutamine-rich tetratricopeptide repeat-containing protein alpha*, *X11Lbeta*, *Transmembrane protein 165*, *Interleukin 16*, *WNT4 protein*, *UNC-112 related protein*,

Table 2

Polymorphic sites of *AmpG* sequences (364 bp) observed in 154 *Rhodnius prolixus* and *R. robustus* I–IV. The column in bold highlights the single-nucleotide polymorphism (SNP) that separates *R. prolixus* from members of the *R. robustus* cryptic species complex (site 280). A dot (.) indicates identity with the consensus nucleotide (in this case, the consensus sequence is from *R. prolixus*). Codes: A – adenine; C – cytosine; G – guanine; T – thymine; S – cytosine or guanine; R – guanine or adenine; K – guanine or thymine; Y – cytosine or thymine.

Species	74	80	167	174	270	280	321	337	341	360
<i>R. prolixus</i>	C	G	G	A	C	A	G	C	C	A
<i>R. robustus</i> I	S	–	–	–	–	G	–	–	–	–
<i>R. robustus</i> II	–	R	R	T	A	G	K	Y	S	T
<i>R. robustus</i> III	–	–	–	T	A	G	T	T	G	T
<i>R. robustus</i> IV	–	–	–	T	A	G	T	–	–	T

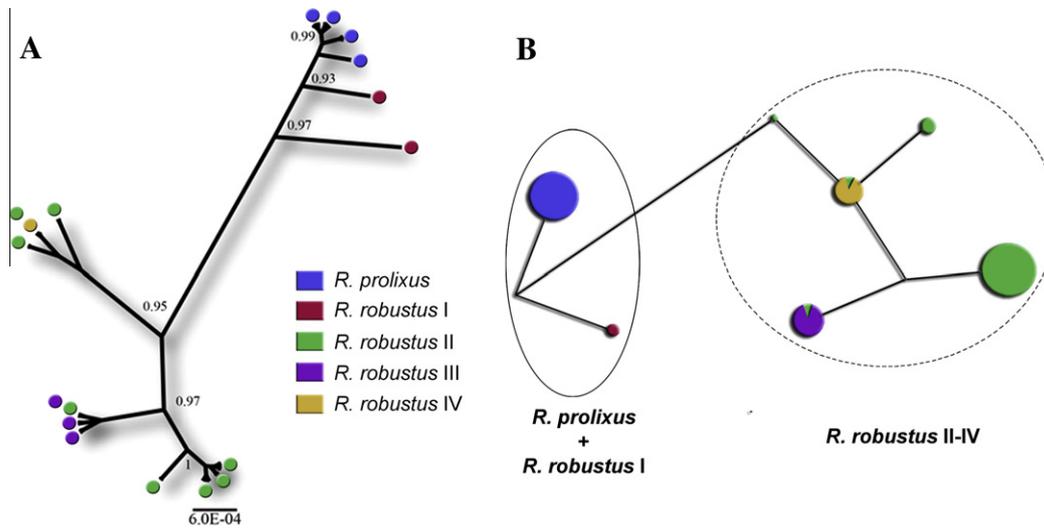


Fig. 2. Phylogenetic relationship between *Rhodnius prolixus* and *R. robustus* I–IV haplotypes. Species are indicated by different colors. (A) Non-rooted 60% majority-rule Bayesian consensus tree obtained from all 154 *AmpG* sequences of *R. prolixus* and *R. robustus*. Posterior probability values higher than 0.9 are indicated near the nodes. Shared genotypes between different *R. robustus* species are represented by more than one circle in a single terminal branch. Note that *R. robustus* I specimens (red circles) are more closely related to *R. prolixus* (blue circles) than to the other *R. robustus* species, evidencing a paraphyletic assemblage. (B) Median-joining network with Maximum-Parsimony post-processing. Each node (or circle) represents a unique haplotype. The size of each node is proportional to the number of specimens that shared the same haplotype, and the branch size is proportional to the number of mutational steps. Note that the distance (measured by branch sizes) between *R. prolixus* and *R. robustus* I is smaller than between *R. robustus* I and *R. robustus* II–IV, confirming their sister-species status. The *R. robustus* paraphyly is shown with the two circles, which highlight the *R. prolixus* + *R. robustus* I group, and the Amazonian *R. robustus* (II–IV), respectively.

Table 3
Maximum-Composite Likelihood (M-CL) estimates of intraspecific (in italics) and interspecific evolutionary divergences distance matrix. The analysis involved all 154 *R. prolixus* and *R. robustus* s.l. *AmpG* sequences. In parentheses, standard deviations calculated after 1000 bootstrap replications. Note that all comparisons between *R. robustus* I and *R. robustus* II–IV resulted in divergence levels higher than between *R. robustus* I and *R. prolixus*.

Species	1	2	3	4	5
1. <i>R. prolixus</i>	<i>0.000</i>				
2. <i>R. robustus</i> I	0.006 (±0.003)	<i>0.005 (±0.003)</i>			
3. <i>R. robustus</i> II	0.012 (±0.007)	0.015 (±0.006)	<i>0.003 (±0.001)</i>		
4. <i>R. robustus</i> III	0.020 (±0.007)	0.017 (±0.007)	0.006 (±0.003)	<i>0.000</i>	
5. <i>R. robustus</i> IV	0.014 (±0.006)	0.011 (±0.004)	0.004 (±0.003)	0.006 (±0.004)	<i>0.000</i>

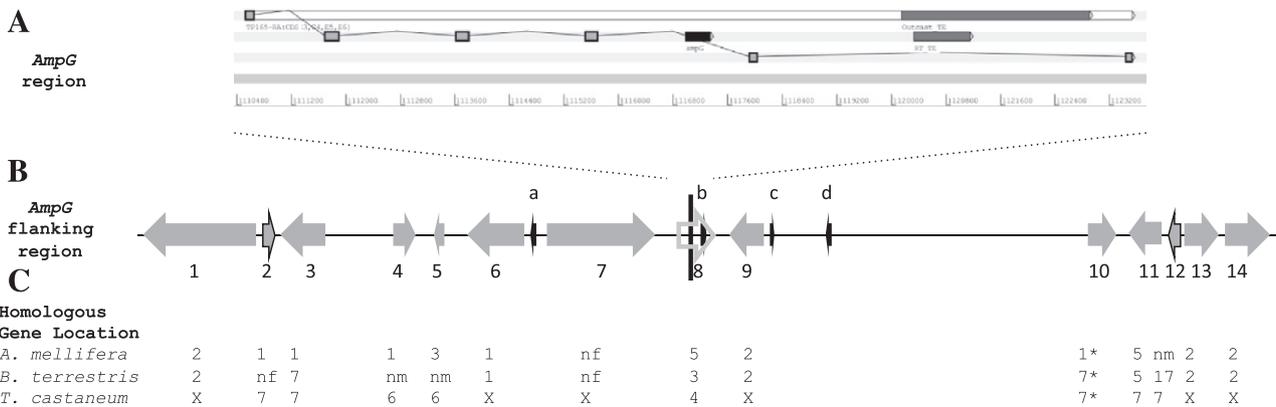


Fig. 3. *AmpG* neighborhood. *Rhodnius prolixus* contig GL563069 (region from 1110523–1123517) that contains *Transmembrane protein 165 (TP165)*. Exons are shown as gray rectangles with black outlines linked together. The *AmpG* region (1116877–1117340) is inside the fourth gene intron. *Outcast_TE* and *RT_TE* are in the fifth intron and represent an outcast class I transposable element and its reverse transcriptase (A). *Rhodnius prolixus* contig GL563069 (region from 925000 to 1319500) contains 14 genes and four transposons. *AmpG* region is shown as a black vertical dashed line. Genes were represented as gray arrows, and EST-supported genes as gray arrows with black outlines. Gene 8 had fully manual curation and EST support, and was represented as a gray outlined arrow (1 – *Ephrin*; 2 – *Ribonuclease*; 3 – *Src tyrosine kinase*; 4 – *Odorant receptor*; 5 – *Sarcoplasmic Ca-binding protein*; 6 – *Small Glu-rich tetratricopeptide repeat-containing protein alpha*; 7 – *X11Lbeta*; 8 – *transmembrane protein 165*; 9 – *interleukin 16*; 10 – *WNT4 protein*; 11 – *UNC-112 related protein*; 12 – *NADH dehydrogenase*; 13 – *KN motif and ankyrin repeat domain-containing protein*; 14 – *band 4.1-like protein 4*). Transposable elements were represented as black arrows ((a) hAT – class II; (b) outcast – class I; (c) mariner – class II; (d) L1 – class I) (B). Homologous genes were identified at the protein level using reciprocal best-hit strategy (Tatusov et al., 1997). Numbers mean chromosome number. X = chromosome; X. nf = homologs not found; nm = homologs not mapped; * = not homologous, only a similar gene (C).

NADH dehydrogenase, KN motif, and ankyrin repeat domain-containing protein and Band 4.1-like protein 4 (Fig. 3B). The detailed information about this region was included in Table A.3.

The *AmpG* region was found in the fourth intron of the *Transmembrane protein 165* gene. This gene's original predictions (RPRC07420-RA and 17948_78) were manually curated (Fig. 3A), including complete EST support. The last intron of this gene also has an *Outcast* transposable element (class I) with a truncated reverse transcriptase (Fig. 3A). Other transposable elements identified were in intergenic regions (Fig. 3B).

3.3. Amplicon G chromosome localization

Arthropod choice for synteny analysis was based on the following criteria: (1) to be the phylogenetically closest organisms with chromosome-mapped genomes available; and (2) to have the closest number of chromosomes to *R. prolixus*, which was known to be 20 autosomes, plus XY (Panzera et al., 2007). *R. prolixus* (taxid 13249) taxonomic classification in the NCBI taxonomic web database (<http://www.ncbi.nlm.nih.gov/taxonomy>) was followed back by tracing the branches of the Insecta phylogenetic tree toward its trunk to search for organisms with chromosome-mapped genomes. Endopterygota Infraclass (taxid 33392), inside Neoptera Subclass (taxid 33340) was the closest taxonomic point that satisfied those two mentioned criteria, where the following 13 species genomes (and their respective number of chromosomes in parenthesis) were found: *A. gambiae* (4), *D. pseudoobscura* (7), *A. mellifera* (16), *Drosophila miranda* (6), *B. terrestris* (18), *M. destructor* (4), *N. vitripennis* (5), *D. virilis* (6), *T. castaneum* (10), *D. simulans* (6), *D. yakuba* (7), *D. melanogaster* (7) and *A. aegypti* (3). Based on the second criterion, we chose for the analysis two bees, *A. mellifera* and *B. terrestris*, and the beetle *T. castaneum*.

Almost all homolog genes of the *AmpG* flanking region were found in the three selected genomes, when we used reciprocal best-hit strategy in gene synteny analysis (Tatusov et al., 1997). The only exception was in the *T. castaneum* analyzed region, where we did not confirm a homolog for the *WNT4* protein (GI 91086553; Fig. 3C), as the best-hit was not reciprocal to *R. prolixus*. However, this protein has the same chromosomal localization as UNC-112 related protein and NADH dehydrogenase (Fig. 3C), and thus was not discarded from analysis. X11Lbeta homolog searches in bees did not retrieve any similar gene, probably due to a lack of prediction, as the TBLASTN web search showed similar genomic regions in unmapped genomic scaffolds (data not shown). *Transmembrane protein 165* gene contained the *AmpG* region inside its fourth intron, but it was the only protein that exhibited homologs located in different chromosomes in all three compared insects (Fig. 3C). Many homolog *R. prolixus* genes were not found, or their chromosome localization did not suggest a pattern in bee genomes, especially *B. terrestris*. Considering *A. mellifera*, four of the first six homolog genes were localized in chromosome 1 (CHR1), but this pattern did not seem to overlap the *AmpG* region. Moreover, no other pattern was observed until the end of the gene block. *T. castaneum* homologs showed a mixed localization pattern in CHRX and CHR7 (six and five occurrences, respectively), but the two homologs flanking *Transmembrane protein 165* gene are located at the X chromosome (Fig. 3C).

Whole contig synteny of contig GL563069 was done with all *T. castaneum* chromosomes separately. Synteny results from the whole contig have confirmed the localization previously identified in reciprocal best-hit approach for the following genes: *Ephrin*, *Src tyrosin kinase*, *Small glutamine-rich tetratricopeptide repeat-containing protein alpha*, *X11Lbeta*, *Transmembrane protein 165*, *Interlukin 16*, *WNT4* protein, *UNC-112 related protein*, *NADH dehydrogenase*, *KN motif* and *ankyrin repeat domain-containing protein* and *Band 4.1-like protein 4* (Fig. A.1). Although the *Ribonuclease* gene did not

show similarity regions neither in CHR7 or CHRX (using an e -value $< 10^{-10}$), its homolog was found in reciprocal best-hit approach (Fig. A.1B). Genes *Odorant receptor* and *Sarcoplasmic calcium-binding protein* were confirmed to be located in CHR6 (data not shown). The overall synteny of this contig in *T. castaneum* CHRX, CHR7 and CHR4 showed 161, 158 and 52 hits, respectively (Fig. A.2) and rates of 14.6, 7.7, and 3.7 hits/Mbase, respectively. The synteny of all other chromosomes presented between 36 and 6 hits and 3.1–0.3 hits/Mbase.

4. Discussion

The *R. prolixus* diagnostic SNP was identified during the initial phase of the *Rhodnius prolixus* genome project, known as *genome survey sequencing* (GSS). GSS sequences will be used, amongst other things, as a framework for the mapping and sequencing of genome size pieces, and this step is required for the assessment of genetic variability values, as high heterozygosity levels could lead to difficulties during genome assembly. This task is accomplished through the identification of variable non-coding single-copy nuclear loci that are then used as markers to estimate colony heterozygosity. In the particular case of the *Rhodnius prolixus* genome project, it was also used with the purpose of identifying a colony that was indeed *R. prolixus*, and not *R. robustus*, or a mixture of the two species. The *amplicon G* (*AmpG*) was the only locus to display a derived character unique to (and thus distinctive of) the *R. prolixus* lineage: an adenine, instead of a guanine, on the 280th position. Due to its usefulness in the colony-screening phase of the project, we decided to evaluate its applicability over a larger sample and geographic area. Our results are very encouraging as they show proven reliability after being tested on an extensive collection of specimens spanning a significant geographical area. Our sampling is evidently not ideal as there are still large areas from where we were unable to obtain specimens. Needless to say, some of these regions are very remote and difficult to access. With respect to the sampling of *R. robustus* I, it is worth mentioning that this species is rare and of restricted geographic distribution. To date, it has only been found in the state of Trujillo, Venezuela.

The ribosomal second internal transcribed spacer (ITS-2) is frequently used for solving taxonomical problems in triatomines. This region seems to be in concerted evolution in some organisms, a molecular process of homogenization between different loci in gene families, as a consequence of gene conversion and unequal recombination (Liao, 1999). This homogenization can be complete, as seems to be the case of *Triatoma* species (Bargues et al., 2002, 2006), or reveal intragenomic and intraspecific variation, as showed in other insects as *Culex* and *Lutzia* species (Diptera: Culicidae) (Vesgueiro et al., 2011). As *Rhodnius* species seem to fit the latter case (CL, MGP, and FAM, unpublished), ITS-2 copies must be cloned and sequenced, which makes its use more expensive and time-consuming.

Although this methodology entails the sequencing of PCR products, as opposed to multiplex-PCR methods (Pavan and Monteiro, 2007), no cloning is required (such as with most rDNA targets). It is also simple and straightforward, because it only involves the identification of a single nucleotide site. We hope that this SNP will be of use for a number of professionals with a broad variety of interests regarding this particular group of insects, however, evaluation of the applicability of this marker over a larger sampling of *R. robustus* I and *R. robustus* IV is desirable. The validation of its potential awaits further usage by the research community that will put it to the test.

In order to determine its genome localization, the *AmpG* region was searched for in all available partial or complete arthropod genomes (40 in total), but it was only located in the genome of *R.*

prolixus itself, in contig GL563069, inside the fourth intron of *Transmembrane protein 165* gene (TP165). Fourteen genes flanking the *AmpG* region were annotated and their homologs searched at the protein level, using reciprocal best-hit strategy (Tatusov et al., 1997) in *A. mellifera*, *B. terrestris* and *T. castaneum*. These were the three arthropods that have shown the closest chromosome number and phylogenetic relationship to *R. prolixus*. Despite the detection of the fourth TPS165 intron as the location of the *AmpG* region, this protein was the only one of the *R. prolixus* contig whose homologs were located on different chromosomes in all three species searched, and thus could not be used to infer *AmpG* localization. Considering all 14 gene homologs' localization in all three species analyzed, *T. castaneum* showed the most homogenous pattern concerning genes location, with six and five occurrences in X and 7 chromosomes (CHRX and CHR7), respectively; notwithstanding, both *AmpG* flanking genes homologs were located in CHRX.

The whole contig synteny approach represents a more reliable method to look at genomic similarities, as DNA sequences are compared along the complete extension of each contig. *R. prolixus* contig GL 563069 synteny in comparison with *T. castaneum* contig also pointed CHRX and CHR7 with almost equal number of hits (161 and 158) as the possible chromosome location, even though their lengths are very different. The frequency of hits in CHRX was two times higher than in CHR7 (14.6 to 7.7 hits/Mbase, respectively) and about ten times higher than all other chromosomes, showing that contig GL563069 shares more potential gene content with *T. castaneum* CHRX. Thus, our results suggest that this contig could be part of *R. prolixus* X chromosome.

DNA sequencing analysis of the *AmpG* region of *R. prolixus* and *R. robustus* *s.l.* samples shows that *R. prolixus* and *R. robustus* I share eight polymorphic sites, three of these are derived nucleotides (A, C, and A, at positions 174, 270, and 360, respectively; Table 2), and hence Maximum-Composite Likelihood divergence between these species reached 0.6% in contrast to 1.1–1.7% between *R. robustus* I and *R. robustus* II–IV. A Bayesian maximum credibility tree and a Median-Joining network with Maximum-Parsimony post-processing further support the paraphyletic nature of *R. robustus* (Fig. 2), previously observed with *cyt b* and rDNA sequences (Monteiro et al., 2003).

In addition to its utility in taxonomy, this marker could be used in combination with the *cyt b* multiplex-PCR (Pavan and Monteiro, 2007) for the detection of natural crossbreeding and introgression of mitochondrial DNA along putative hybrid zones, as reported by Fitzpatrick et al. (2008) in Venezuela. The presence of an adenine/guanine double-peak in electropherograms could be indicative of a recent hybridization event. In addition, as *R. prolixus* has long been used as a model for insect physiology and biochemistry (e.g. Ribeiro et al. (1993) and Wigglesworth (1957)), several laboratories worldwide currently maintain colonies of this species. There is evidence, though, that some of these colonies have been contaminated with, or misidentified as, *R. robustus s.l.* (MGP, CL, EMD, FAM, unpublished). The SNP here described could also be used to evaluate the identity and purity of *R. prolixus* colonies. However, although the possibility of using this SNP as a diagnostic marker for this important vector species seems very promising, its “universal” applicability remains to be evaluated by the research community.

Disclosure statement

The authors declare no conflict of interest.

Role of the funding source

This work was supported by the Brazilian Research Council (CNPq), the Brazilian Ministry of Science, Technology and

Innovation (MCTI), and NIH Grant No. U54 HG003079. The sponsors did not take part nor influenced the any aspect of this study.

Acknowledgements

To John Spieth at the Genome Institute at Washington University School of Medicine in St. Louis for the assistance with library construction and identification of nuclear loci. To Sebastião Aldo Valente, Luis Herman S. Gil, Toby Barrett, José R. Coura, Christine Aznar, and Celia Cordon-Rosales, for kindly providing specimens. We thank José Marcos Ribeiro for providing the Transposon database. The authors are also grateful to the comments of two anonymous referees that helped improve the original manuscript. We also thank the PDTIS-FIOCRUZ DNA sequencing core for running sequencing reactions.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.meegid.2012.10.018>.

References

- Abad-Franch, F., Monteiro, F., 2005. Molecular research and the control of Chagas disease vectors. *An. Acad. Bras. Cienc.* 77, 437–454.
- Abad-Franch, F., Ferraz, G., Campos, C., Palomeque, F.S., Grijalva, M.J., Aguilar, H.M., Miles, M.A., 2010. Modeling disease vector occurrence when detection is imperfect: infestation of Amazonian palm trees by triatomine bugs at three spatial scales. *PLoS Negl. Trop. Dis.* 4, e620.
- Abubucker, S., Martin, J., Yin, Y., Fulton, L., Yang, S.P., Hallsworth-Pepin, K., Johnston, J.S., Hawdon, J., McCarter, J.P., Wilson, R.K., Mitreva, M., 2008. The canine hookworm genome: analysis and classification of *Ancylostoma caninum* survey sequences. *Mol. Biochem. Parasitol.* 157, 187–192.
- Altschul, S.F., Madden, T.L., Schaffer, A.A., Zhang, J., Zhang, Z., Miller, W., Lipman, D.J., 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.* 25, 3389–3402.
- Bandelt, H.J., Forster, P., Rohl, A., 1999. Median-joining networks for inferring intraspecific phylogenies. *Mol. Biol. Evol.* 16, 37–48.
- Bao, Z., Eddy, S.R., 2002. Automated de novo identification of repeat sequence families in sequenced genomes. *Genome Res.* 12, 1269–1276.
- Bargues, M.D., Marcilla, A., Dujardin, J.P., Mas-Coma, S., 2002. Triatomine vectors of *Trypanosoma cruzi*: a molecular perspective based on nuclear ribosomal DNA markers. *Trans. R. Soc. Trop. Med. Hyg.* 96 (Suppl. 1), S159–S164.
- Bargues, M.D., Klisiowicz, D.R., Panzera, F., Noireau, F., Marcilla, A., Perez, R., Rojas, M.G., O'Connor, J.E., Gonzalez-Candela, F., Galvao, C., Jurberg, J., Carcavallo, R.U., Dujardin, J.P., Mas-Coma, S., 2006. Origin and phylogeography of the Chagas disease main vector *Triatoma infestans* based on nuclear rDNA sequences and genome size. *Infect. Genet. Evol.* 6, 46–62.
- Birney, E., Clamp, M., Durbin, R., 2004. GeneWise and Genomewise. *Genome Res.* 14, 988–995.
- Blanco, E., Parra, G., Guigó, R., 2007. Using geneid to identify genes. *Curr. Protoc. Bioinform.* 4, Unit 4.3.
- Coura, J.R., 2007. Chagas disease: what is known and what is needed—a background article. *Mem. Inst. Oswaldo Cruz* 102 (Suppl. 1), 113–122.
- Drummond, A.J., Rambaut, A., 2007. BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evol. Biol.* 7, 214.
- Drummond, A.J., Suchard, M.A., Xie, D., Rambaut, A., 2012. Bayesian Phylogenetics with BEAUti and the BEAST 1.7. *Mol. Biol. Evol.* 29, 1969–1973.
- Fitzpatrick, S., Feliciangeli, M.D., Sanchez-Martin, M.J., Monteiro, F.A., Miles, M.A., 2008. Molecular genetics reveal that silvatic *Rhodnius prolixus* do colonise rural houses. *PLoS Negl. Trop. Dis.* 2, e210.
- Gardner, P.P., Daub, J., Tate, J., Moore, B.L., Osuch, I.H., Griffiths-Jones, S., Finn, R.D., Nawrocki, E.P., Kolbe, D.L., Eddy, S.R., Bateman, A., 2011. Rfam: Wikipedia, clans and the “decimal” release. *Nucleic Acids Res.* 39, D141–145.
- Guhl, F., Pinto, N., Aguilera, G., 2009. Sylvatic triatominae: a new challenge in vector control transmission. *Mem. Inst. Oswaldo Cruz* 104 (Suppl. 1), 71–75.
- Hotez, P.J., Bottazzi, M.E., Franco-Paredes, C., Ault, S.K., Periago, M.R., 2008. The neglected tropical diseases of Latin America and the Caribbean: a review of disease burden and distribution and a roadmap for control and elimination. *PLoS Negl. Trop. Dis.* 2, e300.
- Jukes, T.H., Cantor, C.R., 1969. Evolution of protein molecules. In: Munro, H.N. (Ed.), *Mammalian Protein Metabolism*. Academic Press, New York, pp. 21–123.
- Larkin, M.A., Blackshields, G., Brown, N.P., Chenna, R., McGettigan, P.A., McWilliam, H., Valentin, F., Wallace, I.M., Wilm, A., Lopez, R., Thompson, J.D., Gibson, T.J., Higgins, D.G., 2007. Clustal W and Clustal X version 2.0. *Bioinformatics* 23, 2947–2948.

- Liao, D., 1999. Concerted evolution: molecular mechanism and biological implications. *Am. J. Hum. Genet.* 64, 24–30.
- Lopez, D.C., Jaramillo, C., Guhl, F., 2007. Population structure and genetic variability of *Rhodnius prolixus* (Hemiptera: reduviidae) from different geographic areas of Colombia. *Biomedica* 27 (Suppl. 1), 28–39.
- Marcilla, A., Barges, M.D., Ramsey, J.M., Magallon-Gastelum, E., Salazar-Schettino, P.M., Abad-Franch, F., Dujardin, J.P., Schofield, C.J., Mas-Coma, S., 2001. The ITS-2 of the nuclear rDNA as a molecular marker for populations, species, and phylogenetic relationships in Triatominae (Hemiptera: Reduviidae), vectors of Chagas disease. *Mol. Phylogenet. Evol.* 18, 136–142.
- Monteiro, F.A., Barrett, T.V., Fitzpatrick, S., Cordon-Rosales, C., Feliciangeli, D., Beard, C.B., 2003. Molecular phylogeography of the Amazonian Chagas disease vectors *Rhodnius prolixus* and *R. robustus*. *Mol. Ecol.* 12, 997–1006.
- Panzer, F., Ferrandis, I., Ramsey, J., Salazar-Schettino, P.M., Cabrera, M., Monroy, C., Barges, M.D., Mas-Coma, S., O'Connor, J.E., Angulo, V.M., Jaramillo, N., Perez, R., 2007. Genome size determination in chagas disease transmitting bugs (hemiptera-triatominae) by flow cytometry. *Am. J. Trop. Med. Hyg.* 76, 516–521.
- Pavan, M.G., Monteiro, F.A., 2007. A multiplex PCR assay that separates *Rhodnius prolixus* from members of the *Rhodnius robustus* cryptic species complex (Hemiptera: Reduviidae). *Trop. Med. Int. Health* 12, 751–758.
- Posada, D., 2008. JModelTest: phylogenetic model averaging. *Mol. Biol. Evol.* 25, 1253–1256.
- Ribeiro, J.M., Hazzard, J.M., Nussenzveig, R.H., Champagne, D.E., Walker, F.A., 1993. Reversible binding of nitric oxide by a salivary heme protein from a bloodsucking insect. *Science* 260, 539–541.
- Rutherford, K., Parkhill, J., Crook, J., Horsnell, T., Rice, P., Rajandream, M.A., Barrell, B., 2000. Artemis: sequence visualization and annotation. *Bioinformatics* 16, 944–945.
- Senior, K., 2007. Chagas disease: moving towards global elimination. *Lancet Infect. Dis.* 7, 572.
- Tamura, K., Nei, M., Kumar, S., 2004. Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proc. Natl. Acad. Sci. USA* 101, 11030–11035.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., Kumar, S., 2011. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol. Biol. Evol.* 28, 2731–2739.
- Tatusov, R.L., Koonin, E.V., Lipman, D.J., 1997. A genomic perspective on protein families. *Science* 278, 631–637.
- Tatusova, T., 2010. Genomic databases and resources at the National Center for Biotechnology Information. *Methods Mol. Biol.* 609, 17–44.
- Vesgueiro, F.T., Demari-Silva, B., Malafronte Rdos, S., Sallum, M.A., Marrelli, M.T., 2011. Intragenomic variation in the second internal transcribed spacer of the ribosomal DNA of species of the genera *Culex* and *Lutzia* (Diptera: Culicidae). *Mem. Inst. Oswaldo Cruz* 106, 1–8.
- Wigglesworth, V., 1957. The physiology of insect cuticle. *Annu. Rev. Entomol.* 2, 37–54.